Neural Networks for Secondary Metabolites Prediction in Artemisia Genus (Asteraceae)

Tanja Schwabe, ¹ Marcelo J. P. Ferreira, ¹ Sandra A. V. Alvarenga, ² and Vicente P. Emerenciano. ¹*

Received xxx; Preprint published xxx; Accepted xxx; Published xxx

Internet Electron. J. Mol. Des. 2003, 1, 000–000

Abstract

Motivation. The chemistry of secondary metabolites is a peculiar field of study due to its complexity and the interest it raises in other fields of pharmacology. The plants of the Asteraceae, one of the largest families of plants, have been intensely studied for this reason and have been resulted in the identification of around 28000 occurrences of substances in the species of the family. The chemistry of the Asteraceae is extremely complex and the great problem with databases compiled from the literature is the lack of knowledge about the precision of the data. Thus, the imprecision of the data leads us to use specific techniques to work with this kind of incomplete data. So, the use of artificial neural networks is very adequate. In the present study we focus attention at the genus *Artemisia* and work at the infra genus level in order to try to predict the occurrence of chemical substances present in the genus.

Method. The methodology applied starts by taking all the information on the genus Artemisia from the database. An entry matrix was assembled with the occurrences of the six most representative chemical classes in the genus: flavonoids, monoterpenes, sesquiterpenes, sesquiterpene lactones, polyacetylenes and coumarins. The training of the network was performed with the statistical package $Statsoft^{TM}$ using the backpropagation algorithm.

Results. The secondary metabolites most frequently present in the genus *Artemisia* are monoterpenes and sesquiterpene lactones. Since monoterpenes are present in almost all species, this variable is highly correlated to the variable corresponding of the number total of occurrences. Analyzing the variables corresponding to the sesquiterpene lactones, flavonoids and coumarins show that the two previous ones have similar test set and range errors (c.a. 0.20) while for coumarins, the error is the same, but range falls to half of that.

Conclusions. The results presented show that the mechanism of the neural networks may be effective to predict the occurrence of secondary metabolites in plant genera if an adequate network is used. In this study we show too the application of the artificial neural networks in the chemistry of natural products, a field in which the numerical precision is very small.

Keywords. Artemisia; Asteraceae; Artificial neural networks; secondary metabolites; occurrence prediction.

Abbreviations and notations	
ANN, artificial neural network	MLP, multilayer perceptron
IR, infrared spectra	NMR, nuclear magnetic resonance

1 INTRODUCTION

The chemistry of secondary metabolites is a peculiar field of study due to its complexity and the interest it raises in other fields of pharmacology. This is because of the continuous search for substances with biological activity. On the border between chemistry and biochemistry, the study of new plant species is related to the preservation of the planet's biodiversity, especially in developing

-

¹ Instituto de Química, Universidade de São Paulo, Caixa Postal 26077, 05513-970, São Paulo, SP, Brazil

² Faculdade de Engenharia de Guaratinguetá, Universidade Estadual Paulista, 12516-410, Guaratinguetá, SP, Brazil

^{*} Correspondence author; E-mail: vdpemere@iq.usp.br; phone: 55-11-3091-2056; fax: 55-11-3815-5579;

countries where the destruction of the forest takes place at alarming rates.

Compiling data on secondary metabolites already isolated from plants, thus "transforming" these into a database, is a hard task which our research group has undertaken. We have centered our efforts on the Asteraceae, one of the one of the largest families of plants [1,2].

The chemistry of the Asteraceae is extremely complex, in accordance with of its biological and anatomical complexity, based on morphological criteria [3] and DNA variation [4]. Several research groups have dedicated efforts to the study of Asteraceae plants in the laboratory. Among these, the group of Bohlmann [5] stands out. At that time, about 7,000 compounds had been isolated from plants of this family. Large reviews have been produced on some classes of secondary metabolites in the Asteraceae. Sesquiterpene lactones were reviewed by Seaman [6], diterpenes were reviewed by Bohlmann et al.[7], flavonoids by Emerenciano [8] and Bohm [9], triterpenes by Macari [10] polyacetylenes by Bohlmann [11], benzofuranes and benzopiranes by Proksck [12]. All these data were updated and used in an analysis of the subgroups of the family, using PCA techniques [13].

The great problem with databases compiled from the literature is the lack of knowledge about the precision of the data. This is because these data are subjected to laboratory errors, studies directed to the search for a specific chemical class, etc. There also the implications of geographical and ecological factors, as well as the phenotypic pressures all influencing the production of secondary metabolites. Thus, the imprecision of the data leads us to use specific techniques to work with this kind of incomplete data.

The use of neural networks has become a routine in chemistry. A review by Zupan and Gasteiger [14] describes some of the applications of neural networks in chemistry and later the examples presented in this review were enhanced in a book [15]. Virtually all areas of chemistry have scientific studies published using neural networks. It is interesting to note the applications on ¹³C and ¹H NMR [16,17], in IR and mass spectra [18,19]. Applications of neural networks in natural products chemistry are somewhat less frequent. The study of oxidized triterpenes or limonoids by Fraser et al. [20] provides an example. In chemical taxonomy our groups is working with a large database for the prediction of the occurrence of substances in genus of Asteraceae [21]. In the study mentioned we were able to predict the presence of chemical classes in ca. 500 genera of Asteraceae.

In the present study we focus attention at the genus *Artemisia*. We work at the infra genus level in order to try to predict the occurrence of chemical substances present in the genus.

2 METHODS

The methodology applied starts by taking all the information on the genus *Artemisia* from the database. According to Ling [22,23] there are ca. 400 described species of *Artemisia* in the world and our database contains chemical data on about half of these which is fundamental for the training

of the network.

An entry matrix was assembled with the occurrences of the six most representative chemical classes in the genus: monoterpenes, sesquiterpenes, sesquiterpene lactones, flavonoids, polyacetylenes and coumarins. Figure 1 describes a typical substance from each class of compounds analyzed in this study [24-27]. We define occurrences as the number of times a substance of a certain class was present in a taxon. For example, if the database contains for two different species of the same tribe i n_1 compounds belonging to the chemical class j for the first species and n_2 compounds of the same chemical class for the second species, the number of occurrences $O_{i,j}$ in the tribe i is the sum corresponding to $n_1 + n_2$. If a unique compound is isolated from two different species of the same tribe, it is counted twice.

Figure 1. Typical compounds from each class of secondary metabolites analyzed in this study

As the number of occurrences may be dependent on the factors mentioned above, we introduced other variables in the matrix to describe the presence or absence of the chemical classes in each case analyzed. These variables, referred to as heuristic, describe the probability of a certain species to present or not a certain chemical class. We defined arbitrarily the presence of a chemical class in a species as positive (1.0) if the value of the continuous variable of that species is greater than or equal to the average of all cases. Otherwise the heuristic variable equals zero. The twelve variables used as inputs for the network are described in table 1.

The training of the network was performed with the statistical package Statsoft™ [28] using the backpropagation algorithm and adapting the training time for the each type of variable. For the training set we used 180 cases and we used 40 cases for the test set. Training time in a Pentium Microcomputer with a clock of 1 GHz varied from 5 minutes to 5 hours as described in table 2. The structure of the best network chosen was of the type MLP 11:11-7-1:1 and 10:5:1:1.

Table 1. Variables used as inputs for the network

Mono = Monoterpenes

Sesq = Sesquiterpenes

Lact = Sesquiterpene lactones

Cuma = Coumarins

Poli = Polyacetylenes

Flav = Flavonoids

CLM = Presence or absence of Monoterpenes

CLS = Presence or absence of Sesquiterpenes

CLL = Presence or absence of Sesquiterpene lactones

CLC = Presence or absence of Coumarins

CLP = Presence or absence of Polyacetylenes

CLF = Presence or absence of Flavonoids

Total = Sum of occurrences in the taxon

NCLA = Sum of the number of classes in the taxon

In total, 3200 networks were retained after several days of training. Table 2 describes only the best network obtained using the "custom design" option. The networks obtained had to be retrained ca. 10 times using the following procedure: 10000 = epochs, learning rate = 0.01, momentum = 0.3. The results of the network were compared with the descriptive statistics for the variables and with a correlation matrix for the variables (Table 2).

Table 2. Results obtained from the best neural network

			Standard.		Train	Test	Train	Test	Net
Variable	Range	Mean		Variance					
			Deviation		Error	Error	Perform.	Perform.	Structure
Mono	171	10.105	21.756	473.312	0.244	0.411	0.999	0.999	9:9-8-1:1
Sesq	13	0.405	1.693	2.877	0.338	0.817	0.945	0.980	9:9-8-1:2
Lact	44	3.642	7.180	51.564	1.795	4.591	0.969	0.940	12:12-3-1:1
Cuma	10	0.819	1.893	3.583	0.008	0.011	0.999	0.949	13:13-1-1:1
Flav	37	1.374	4.533	20.553	0.029	0.146	0.999	0.948	13:13-1-1:1
Poli	13	0.879	2.187	4.784	0.218	0.511	0.976	0.964	13:13-1-1:1
CLM	1	0.368	0.484	0.234	0.120	0.141	0.929	0.920	13:13-1-1:1
CLS	1	0.084	0.278	0.077	0.140	0.108	0.939	0.917	13:13-10-1:1
CLL	1	0.310	0.464	0.215	0.175	0.230	0.836	0.786	13:13-4-1:1
CLC	1	0.242	0.429	0.185	0.128	0.259	0.919	0.840	13:13-4-1:1
CLF	1	0.126	0.333	0.111	0.104	0.092	0.895	0.855	13:13-2-1:1
CLP	1	0.226	0.419	0.176	0.118	0.276	0.956	0.807	13:13-18-1:1
NCLA	5	1.798	1.022	1.022	0.274	0.362	0.809	0.862	13:13-18-1:1
TOTAL	227	17.216	29.028	29.028	0.374	0.476	0.999	0.999	13:13-4-1-:1

3 RESULTS AND DISCUSSION

It is adequate to inspect the data obtained both from the chemical and the computational points of view. The secondary metabolites most frequently present in the genus *Artemisia* are monoterpenes and sesquiterpene lactones featuring 171 and 35 occurrences, and flavonoids (range = 37). The data in Table 3 show a great variance for these data when compared with the other variables. These are also the variables where the errors of the network were the smallest both in the test set and in the training set if we take them as percentages. Obviously, the total number of substances in a certain species may also be large, if there are more studies on it in the literature. Since monoterpenes are present in almost all species, this variable (MONO) is highly correlated to the variable TOTAL.

Table 3. Correlation matrix of the variables in *Artemisia* genus

	Mono	Sesq	Lact	Cuma	Flav	Poli	CLM	CLS	CLL	CLC	CLF	CLP	NCLA	TOTAL
Mono	1.00	0.35	0.15	0.32	0.45	0.27	0.55	0.26	0.08	0.09	0.19	0.00	0.44	0.52
Sesq	0.35	1.00	0.14	0.08	0.35	0.01	0.31	0.79	0.06	-0.04	0.12	0.00	0.41	0.44
Lact	0.15	0.14	1.00	-0.12	0.03	0.26	0.08	0.27	0.72	-0.14	-0.03	0.00	0.33	0.41
Cuma	0.32	0.08	-0.12	1.00	0.27	0.09	0.08	0.05	-0.19	0.71	0.06	0.00	0.33	0.34
Flav	0.45	0.35	0.03	0.27	1.00	-0.03	0.09	0.25	0.00	0.12	0.68	0.00	0.37	0.43
Poli	0.27	0.01	0.26	0.09	-0.03	1.00	0.15	0.14	0.12	0.05	0.00	0.00	0.40	0.20
CLM	0.55	0.31	0.08	0.08	0.09	0.15	1.00	0.40	-0.02	-0.03	0.01	0.00	0.37	0.61
CLS	0.26	0.79	0.27	0.05	0.25	0.14	0.40	1.00	0.13	-0.03	0.10	0.00	0.41	0.57
CLL	0.08	0.06	0.72	-0.19	0.00	0.12	-0.02	0.13	1.00	-0.23	-0.02	0.00	0.26	0.41
CLC	0.09	-0.04	-0.14	0.71	0.12	0.05	-0.03	-0.03	-0.23	1.00	0.00	0.00	0.26	0.34
CLF	0.19	0.12	-0.03	0.06	0.68	0.00	0.01	0.10	-0.02	0.00	1.00	0.00	0.31	0.39
CLP	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	1.00	0.00	0.00
NCLA	0.44	0.41	0.33	0.33	0.37	0.40	0.37	0.41	0.26	0.26	0.31	0.00	1.00	0.68
TOTAL	0.52	0.44	0.41	0.34	0.43	0.20	0.61	0.57	0.41	0.34	0.39	0.00	0.68	1.00

Obviously, the most frequent secondary metabolites in the database are those most isolated by researchers and this number is debatable from several points of view. From the laboratory point of view, we can mention the throwing away of extract fractions *a priori* considered unproductive by the researcher. From the analytical point of view there are the difficulties of interpretation of spectral data (every time more improbable, since the appearance of 2D NMR techniques). From the phenotypic point of view, it is accepted among phytochemists that the production of secondary metabolites is connected to the attraction or repulsion due to interactions among plants, among plants and insects, the soil, etc, and differences among genotypes. Although we work in a low hierarchical level (genus) the taxonomic units not always constitute natural groups of species. These are also subjected to erroneous classifications and it is thus impossible to warrant the presence of a species in the same genus until studies of molecular biology [29] corroborate the morphological data. Lastly, the database is not complete because some isolated compounds may not have been

published or the collect of data in the literature may not have been precise. Therefore, a small database such as the one cited here, with imprecise information becomes adequate to an inspection *via* neural networks.

The errors observed when one works with gross number of occurrences must be regarded carefully in function of the factors mentioned. As an example of this, a simple inspection of the variables LACT, FLAV CUMA and TOTAL in Tables 1 and 2 show that the two previous ones have similar test set and range errors (c.a. 0.20) while for CUMA, the error is the same, but range falls to half of that. The TOTAL variable the error for the test set is 0.13 and it includes all other variables

The use of heuristic variables (presence/absence) based on the average of every variable was adopted here to try to introduce variables that escape the mentioned sources of error.

4 CONCLUSIONS

The results presented in Table 2 show that the mechanism of the neural networks may be effective to predict the occurrence of secondary metabolites in plant genera if an adequate network is used.

In this study we show the application of the ANN in the Chemistry of Natural Products, a field in which the numerical precision is very small.

It was also demonstrated that the problem of the predictability may be treated through the ANN in a vast universe such as phytochemistry. As one works in very low hierarchical levels in the botanical point of view, the errors in the data may be even greater. But even so, the mechanism of neural networks, and, more precisely, the adequate training with the best cases of the available sample, were able to produce results that may be considered good. It was also observed that, in spite of the strong ability to deal with imprecise data, the skill of the chemist to analyze the first training sets was paramount for the performance of the ANN.

The use of mathematical models in biology is not new, but models that are general and can produce good predictions will be useful to direct, or at least orient, laboratory work in search of new active principles of plants.

Acknowledgment

This work was supported by grants from the Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) and by the Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq).

5 REFERENCES

[1] K. Bremer, Asteraceae. Cladistics and Classification, Timber Press, Portland, Oregon, 1994.

- [2] K. Bremer, Major clades and grades of the Asteraceae; in: Proceedings of the International Compositae Conference, Eds. D. J. N. Hind and H. J. Beentje, Royal Botanic Gardens, Kew, 1996, *I*, Systematics, pp. 1-7.
- [3] K. R. Sporne, Ovule as an indicator of evolutionary status in angiosperms, New Phytol. 1969, 68, 555-566.
- [4] R. K. Jansen, K. E. Holsinger, H. J. Michaels and J. D. Palmer, Phylogenetic analysis of chloroplast DNA restriction site data at higher taxonomic levels: an example from the Asteraceae, *Evolution* **1990**, *44*, 2089-2105.
- [5] C. Zdero and F. Bohlmann, Systematics and evolution within the Compositae, seen with the eyes of a chemist, *Pl. Syst. Evol.* **1990**, *171*, 1-14.
- [6] F. Seaman, Sesquiterpene lactones as taxonomic markers in the Asteraceae, Bot. Rev. 1982, 48, 121-595.
- [7] F. Seaman, F. Bohlmann, C. Zdero and T. J. Mabry, *Diterpenes in flowering plants: Compositae (Asteraceae)*, Springer, New York, 1990.
- [8] V. P. Emerenciano, M. A. C. Kaplan, O. R. Gottlieb, Evolution of sesquiterpene lactones in angiosperms, *Biochem. Syst. Ecol.* **1985**, *13*, 145-166.
- [9] B. A. Bohm and T. F. Stuessy, Flavonoids of the sunflower family, Springer-Wien, New York, 2001
- [10] P. A. T. Macari, J. P. Gastmans, G. V. Rodriguez and V. P. Emerenciano, An expert system for structure elucidation of triterpenes, *Spectroscopy- An International Journal* **1994/1995**, *12*, 139-166.
- [11] F. Bohlmann, T. Burkhardt and C. Zdero, Naturally occurring acetylenes, Academic Press, London, 1973.
- [12] P. Proksch and E. Rodriguez, Chromenes and benzofuranes of the Asteraceae, their chemistry and biological significance, *Phytochemistry* **1983**, *22*, 2335-2355.
- [13] S. A. V. Alvarenga, M. J. P. Ferreira, V. P. Emerenciano and D. Cabrol-Bass, Chemosystematic studies of natural compounds isolated from Asteraceae. Characterization of tribes by principal component analysis, *Chem. Intell. Lab. Syst.* **2001**, *56*, 27-37.
- [14] J. Gasteiger and E. Zupan, Neural networks in Chemistry. Angew. Chem. Int. Ed. Engl. 1993, 32, 503-527.
- [15] E. Zupan and J. Gasteiger, Neural Networks for Chemists An Introduction, VHC-Verlag, Weinhein, 1993.
- [16] J. Doucet, A. Panaye, E. Feuilleaubois and P. Ladd, Neural networks and ¹³C NMR chemical shift prediction, *J. Chem. Inf. Comput. Sci.* **1993**, *33*, 320-324.
- [17] J. Aires-de-Souza, M. C. Hemmer and J. Gasteiger, Prediction of ¹H NMR chemical shifts using neural networks, *Anal. Chem.* **2002**, *74*, 80-90.
- [18] C. Cleva, C. Cachet and D. Cabrol-Bass, Clustering of infrared spectra with Kohonen networks. *Analusis* **1999**, 27, 81-90.
- [19] H. Lohninger and F. Stanci, Comparing the performance of neural networks to well-established methods of multivariate data analysis: the classification of mass spectral data, *Fresenius J. Anal. Chem.* **1992**, *344*, 188-189.
- [20] L. Fraser, D. A. Mulholland and D. D. Fraser, Classification of limonoids and protolimonoids using neural networks. *Phytochem. Anal.* **1997**, *8*, 301-311.
- [21] M. J. P. Ferreira, A. J. C. Brant, S. A. V. Alvarenga, F. M. M. Magri, A. R. Rufino and V. P. Emerenciano, Prediction of occurrences of diverse chemical classes in Asteraceae through neural networks, *Phytochem. Anal.* **2003**, in press.
- [22] Y. R. Ling, The Old World Artemisia Linn. (Compositae). Bull. Bot. Res. Harbin 1992, 12, 1-108.
- [23] Y. R. Ling, The New World *Artemisia*; in: *Advances in Compositae Systematics*, Eds. D. J. N. Hind, C. Jeffrey and G. V. Pope, Royal Botanic Gardens, Kew, 1995, pp. 255-281.
- [24] G. D. Brown, G. -Y. Liang and L. -K. Sy, Terpenoids from the seeds of *Artemisia annua*, *Phytochemistry* **2003**, 64, 303-323.
- [25] R. X. Tan, H. Q. Tang, J. Hu and B. Shuai, Lignans and sesquiterpene lactones from *Artemisia sieversiana* and *Inula racemosa*, *Phytochemistry* **1998**, *49*, 157-161.
- [26] T. –S. Wu, Z. –J. Tsang, P. –L. Wu, F. –W. Lin, C. –Y. Li, C. –M. Teng and K. –H. Lee, New constituents and antiplatelet aggregation and anti-HIV principles of *Artemisia capillaris*, *Bioorg. Med. Chem.* **2001**, *9*, 77-83.
- [27] J. A. Marco, J. F. Sanz-Cervera, F. Sancenón, M. Arnó and J. Vallès-Xirau, Sesquiterpene lactones and acetylenes from *Artemisia reptans*, *Phytochemistry* **1994**, *37*, 1095-1099.
- [28] Statistica Neural Networks, Statsoft, Inc. Tulsa, OK, EUA, 2000.
- [29] E. E. Schilling and J. L. Panero, A revised classification of subtribe Helianthinae (Asteraceae: Heliantheae). I. Basal lineages, *Bot. J. Linn. Soc.* **2002**, *140*, 65-76.